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Blowflies Reared in Laboratory Conditions from Maggots Collected on Rat (*Rattus norvegicus* Berkenhout, 1769, Var Wistar) Carrions in Yaoundé (Cameroon, Central Africa)

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Authors' contributions

This work was carried out in collaboration among all authors. Author FDFY designed the study, perform the field work, the laboratory survey, the writing of manuscript and the identification of specimens, author PBN participated to the field work and the laboratory survey, authors YB and MHV conceptualize, revise the identification and performed the litterature revue, author CD-I managed the data analysis and author CFBB supervise the field work, data analysis and writing and revising of manuscript. All authors read and approved the final manuscript.

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Original Research Article

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ABSTRACT

Forensic entomology offers insects as physical evidence during legal procedures. Forensic entomologists have determined succession of arthropods on dead animals, but few published studies are available on necrophagous larvae collected on carcasses around the world. This study evaluated the diversity of arthropods associated with rat carcasses to identify species of forensic relevance. Larvae hatched from arthropod eggs were reared until the emergence of adult flies under ambient laboratory conditions. Adult flies were identified to species level. Overall, 6319 individuals belonging to 6 families, 13 genera and 21 species of Diptera emerged.

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1. INTRODUCTION

In the course of legal procedures, investigators are expected to produce evidence during inquiries at the court. Nowadays, it is a difficult task to produce palpable evidence without the intervention of scientists such as forensic entomologists, toxicologists, ballistic experts, etc. [1]. One milestones is to determine time elapsed since death, known as Post Mortem Interval (PMI), and the circumstances of the death. Judicial investigators need accurate data that help them to make decisions. Within three days (72 hours) after death, legal medicine can use autopsy to obtain data for the court [2]. After this time, accurate data can be obtained from studies targeting on necrophagous arthropods sampled on and around the corpse [3,4,5].

Forensic, judicial or medico-legal entomology is the science that deals with the study of necrophagous arthropods collected at crime scenes and assessed to inform cases in court [1]. The principle of predictable species succession on corpses usable in judicial inquiries was recognized by Megnin [6] who suggested eight successive waves of arthropods colonizing a carrion over the time. This principle has been debated because the delimitation of the different waves is a difficult task [7-11]. The time of arrival of a specific insect on the cadaver can be monitored according to biogeographic variation [12]. To gather data for each environment, it is crucial to conduct local studies and determine the insect species involved in decaying corpses [10].

Africa is the stronghold of over a dozen necrophagous blowfly species, several of which are found on other continents [13]. These flies have been intensively studied over the world, but papers referring to African countries are scarce [14] or absent, except in South Africa [15,4,10,14,16], Nigeria [17,18], Namibia [19], Algeria [20-25] and Cameroon [26,27]. Consequently, crime cases solved with entomological evidence and exploitation of the ecological database are too poor or remain scarce in Africa because (1) a lot of spiritual value is attached to corpses, (2) some investigators or researchers do not know enough about forensic entomology and. (3) judicial employees rarely ask for scientific or palpable proof during their procedures [1,28].

The aim of this study is to identify and document all forensic relevant blowflies by rearing larvae hatched from eggs until the emergence of adults under laboratory conditions.

2. MATERIALS AND METHODS

2.1 Study Site

This study was conducted in an experimental station located on the premises of the University of Yaounde I (11°33'01″E 3°51'35″N), Cameroon. The climate is equatorial and characterized by four distinct seasons: a long rainy season from mid-November to February, a short rainy season from March to June, a short dry season from July to August and a long rainy season from September to mid-November. The annual mean temperature is about 23-24°C and the rainfall fluctuates between 1500 and 2000 mm [29,30]. The landscape of this part of the campus suburban and characterized by the presence some trees like *Elaeis guineensis* (Arecaceae) and *Musa* sp. (Musaceae).

2.2 Experimental Procedure

At the experimental station, two rats Rattus norvegicus Berkenhout, 1769, var Wistar were euthanized with carbon dioxide and then strangulated. These two carcases were immediately deposited on a 10 cm layer of sterilized soil inside a rectangular (40 cm x plastic container. 20 cm) These two carcasses were protected against animal scavengers with a wooden cage (120 cm x 120 cm x 120 cm) covered with a net of 5 cm mesh size to allow colonization of the carcasses by insects only. The cage had an entrance allowing visiting and checking for larvae migrating to the sterilized soil. At the end of each search, the remains were again placed on top of fresh sterilized soil. The collected larvae and the cadaveric liquid were covered with a net to avoid eventual contamination by subsequent insect visitors and taken to the laboratory for rearing to maturity under ambient air conditions.

The plastic box was inspected twice daily, at 08:00 GMT and 16:00 GMT, to record the emergence of adult flies. The litter was immediately transferred to another plastic box and the adult flies were reared and fed for 48 hours with honey put on cotton and placed inside the box [31,32]. After this period of time,

these adult flies were captured using flexible plastic forceps and preserved in 70% ethanol in microreaction tubes for further morphological identification. This exercise was repeated for five days and stopped when no new emergence was noticed.

Atmospheric temperature and relative humidity were recorded using a data logger Testo.

2.3 Fly Identification

The identification of adult flies was performed in two phases. The first phase, done using a stereomicroscope (M3Z,Herbrugg Switzerland), wa based on identification keys [33-36] [19,37,38,26,39,13]. The second step was performed successively at the Microtraces Laboratory of the National Institute of Criminalistic and Criminology of Brussel and the entomology section of the laboratory of Royal Museum of Central Africa at Tervuren, all in Belgium. It used the identification keys of Kurahashi & Chowanadisal [40], Rognes & Paterson [41], Kurahashi & Kirk-Spriggs [19], Couri [42], Carvalho & Mello-Patiu [43], Nihei & Carvalho [38] and LinLong et al. [44].

2.4 Data Analysis

The rate of occurrence (C) of each species was evaluated according to the formula [45] C = (p*100)/P, where p is the number of occurrences of a given species during the survey and P = 5 i. e. the total number of replicates. The results are interpreted as C > 50% = consistant species; $25\% \le C \le 50\%$ = accessory species; C < 25\% = rare species.

3. RESULTS

3.1 Abiotic Parameters

During the experiment, the mean daily temperature was 28.5°C (21.4-33.5°C) and the mean daily relative humidity was 64.65% (56.9-85.1%).

3.2 Overview of Emerged Entomofauna

The entomofauna that emerged was made up of 6 families, 13 genera and 21 species. Amongst the families, the Braconidae, Diapriidae and Pteromalidae are parasitoid wasps. Braconidae predominated among them and were present at each trial. When any of these families occurred together, the decreasing order of abundance was always Braconidae > Diapriidae > Pteromalidae (Table 1). With the exception of the first rearing experiment where only 7 species emerged, during the other trials 16 to 19 species emerged. The decreasing order of abundance was Calliphoridae, Muscidae, Braconidae, Sarcophagidae, Diapriidae and Pteromalidae (Table 1).

During the first trial, Calliphoridae, Muscidae, Sarcophagidae and Braconidae represented 42.85%, 37.71%, 11.9%, and 9.52% of the entomofauna. The two main species were *Hydrotaea* sp. (35.71%) and *Lucilia* sp. (28.57%) (Fig. 1).

During the second trial, the frequencies of insects were 84.72%, 12.9%, 2.02%, and 0.4% for Calliphoridae, Muscidae, Sarcophagidae and Braconidae respectively. The main species were Calliphoridae, namely *Hemipyrellia fernandica* (40.03%), *Chrysomya putoria* (19.50%) and *C. albiceps* (14.49%) (Fig. 1).

During the third trial, the entomofauna was made up of Calliphoridae (84.40%), Muscidae (7.50%), Braconidae (4.50%), Diapriidae (2.3%), Sarcophagidae (1.2%) and Pteromalidae (0.1%). The main species were *Hemipyrellia fernandica* (43.60%), and *C. putoria* (20.07%) (Calliphoridae) (Fig. 1).

The fourth tiral made it possible to obtain Calliphoridae (82.50%), Muscidae (16.27%), Braconidae (0.5%), Sarcophagidae (0.41%) and Pteromalidae (0.20%). The main species were *Hemipyrellia fernandica* (46.07%), *Chrysomya putoria* (15.86%) and *Chrysomya* sp. (11.00%) (Fig. 1).

At last during the fifth trial, insects that emerged were Calliphoridae (90.50%), Muscidae (6.60%), Braconidae (2.15%), Sarcophagidae (0.9%), Diapriidae (0.50%) and Pteromalidae (0.20%). The main species were *Hemipyrellia fernandica* (58.65%) and *Chrysomya putoria* (15.93%) (Fig. 1).

During the successive trials, only three (Calliphoridae, Muscidae families and Sarcophagidae) constituted the dominant necrophagous entomofauna. Calliphoridae predominated and represented 47.40% (18 individuals), 86.5% (639 individuals), 90.70% (1188 individuals), 83.20% (1596 individuals) and 93. 20% (1971 individuals) amongst that necrophagous entomofauna (Table 1). As far as necrophagous arthropods are concerned, almost all species were constant (C > 50%), except Sarcophaga zumpti, which was accessories (C =40%).

4. DISCUSSION

Several works have reported faunistic inventories of necrophagous insects over the world, but data on emerged adult flies from larvae collected on cadavers are scarces [10]. The current work is the first initiative in Central Africa to rear larvae during forensic а entomology experiment within ambient laboratory conditions.

4.1 Carrion Models

A variety of baits are used for forensic purposes. In the present laboratory study, rats were used as trapping bait like in other experiments already documented [46,20,21,9,2,47,48,49]. [50-53] used carcasses of pigs as trapping substrates. [54,55,56,12,57] utilized multiple trapping baits while [58,59] employed fish when [60,61] used chicken liver as bait. This multitude of biological models of trapping substrates gives new opportunities to authors to enrich their database on forensic entomology.

4.2 Emerged Necrophagous and Parasitoid Entomofauna

The present survey revealed 6 families, 13 genera and 21 species. This entomofauna diversity is greater than that published by [62-67]. These scholars censured twelve to fourteen species excluding beetles within their differences in climatic conditions and local arthropod assemblages since their studies were carried out in Europe, Asia, North America and Australia respectively. This supports the fact that the biodiversity of these insect is governed by the environmental parameters specific to each biogeographic area [28,68] and also the body size of the bait.



Fig. 1. Evolution of the entomofauna that emerged in the different trial

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	Trial							Total per family						
Таха	1		2		3	3		4		5				
	n	%	n	%	n	%	n	%	n	%	Species	%	Family	%
DIPTERA														
Calliphoridae														
C. albiceps			10 9	14.4 9	109	7.70	77	3.98	81	3.71	376	5.95		
C. laxifrons			3	0.40	8	0.57	2	0.10			13	0.21		
Chrysomya putoria	2	4.76	14 7	18.2	284	20.0 7	307	15.8	348	15.9 3	1078	17.0		
Chrysomya sp.			19	2.53	78	5.51	213	11.0 0	119	5.45	429	6.79	5402	85.4
Hemipyrellia fernandica	4	9.52	30 1	40.0 3	617	43.6 0	892	46.0 7	128 1	58.6 5	3095	48.9 8	5102	9
Hemipyrellia sp.			22	2.93	40	2.83	48	2.48	57	2.61	167	2.64		
Lucilia cuprina							5	0.26	2	0.09	7	0.11		
Lucilia sp.	1 2	28.5 7	38	5.05	52	3.67	52	2.69	83	3.80	237	3.75		
Muscidae														
Atherigona sp.			16	2.13	15	1.06	1	0.05	31	1.42	63	1.00		
<i>Hydrotaea</i> sp.	1 5	35.7 1	46	6.12	58	4.10	98	5.06	42	1.92	249	3.94	665	10.5
<i>Musca</i> sp.			25	3.32	23	1.63	152	7.85	49	2.24	249	3.94		2
<i>Ophyra</i> sp.			10	1.33	9	0.64	64	3.31	21	0.96	104	1.65		
Sarcophagidae														
S. africa			8	1.06	7	0.49	2	0.10	5	0.23	22	0.35		
Sarcophaga sp.	4	9.52	3	0.40	12	0.85	8	0.41	1	0.05	28	0.44	56	0.89
Sarcophaga zumpti HYMENOPTERA	1	2.38			5	0.35					6	0.09		
Braconidae														
Apanteles sp.			6	0.80			1	0.05	1	0.05	8	0.13		
Coelalysia nigriceps	4	9.52	8	1.06	45	3.18	10	0.52	47	2.15	114	1.80		
Coelalysia sp.					13	0.92	1	0.05	2	0.09	16	0.25	139	2.20
<i>Undetermined</i> (genera and species)			1	0.13	6	0.42					1	0.02		
Trichopria sp.					33	2.33			10	0.46	43	0.68	43	0.68
Pteromalidae														
Spalangia sp.						100								
Total per trial	4	100	75	100	141	2	193	100	218 2	100	6319	100	6319	100

Table 1. Number (n) and proportion (%) of emerged flies in the different trial

Six families (Table 1) were obtained in this work, whereas [67] collected individuals belonging to Fanniidae, Piophilidae, Sepsidae, Sphaeroceridae, Phoridae, and Heleomyzidae during their experiment in Central Quebec, Canada, an environment dominated by a snowy winters and humid and temperate summers [69]. This suggests that type of vegetation and abiotic conditions are determinant factors during forensic entomology studies.

Our experiment showed the predominance of Calliphoridae (*Lucilia* sp., *Lucilia* cuprina, *Chrysomya albiceps*, *C. putoria*, *C. laxifrons*, *Hemipyrellia fernandica*, *Hemipyrellia* sp. and *Chrysomya* sp). This result confirms the work of Ekenem & Dike [17] in Nigeria, despite the fairly differences in the genera/species richness, and also corroborates the results of Matuszewski et al. [70,71] in Central Europe, Fremdt and Amendt [72], Bernhardt et al. [73] in Germany, and Thyssen et al. [54] in Brazil.

Muscidae were the second-most prevalent family in the current work, as observed in other studies performed in other biogeographic areas. In fact, [52,74] also reported the presence of Muscidae during a forensic set up. The Muscidae species that emerged in the present work are different from those obtained by Pitner et al. [75] in northern Germany. This could be the effect of environmental parameters which guide the geographical distribution of necrophagous flies. through their thermophysiological tolerances [76,77]. entomology Consequently, forensic experiments should be carried out in every region due to biogeographical specificity. This differences might also be due to vegetation, phylogenetic history or even season.

Sarcophagidae also emerged in our trials. That was the case in South Africa [78,10], Central Spain [79], Iran [80-82], central Quebec at Canada [67], and Nigeria [17]. The difference in species diversity may be the result of differences of geographic location and environmental factors (temperature, photoperiod, light, water or moisture and relative humidity), since the biodiversity and abundance of necrophagous flies are influenced by these abiotic parameters [28].

Braconidae, Pteromalidae and Diapriidae emerged during the present rearing experiments contrary to the results reviewed by Villet [10,81]. The genera *Nasonia* (= *Mormoniella*) on the one hand, and *Nasonia* and *Brachymeria* on the other, were obtained in South Africa [10] and Iran [81], respectively. Grassberger & Franck [83] in Punjabi India, Turchetto et al. [84] in Italy and Voss et al. [85] in Western Australia, collected other different parasitoid wasps during their studies. In a similar experimental design by Marchiori [86] and Horenstein & Salvo [86] in Brazil, Pteromalidae species also emerged from reared Calliphoridae larvae; this suggests that wasps of this taxon also infest necrophagous larvae worldwide. entomofauna In our environment, Braconidae may occur alone or be associated with Diapriidae and/or Pteromalidae: the two latter families being weakly represented.

5. CONCLUSION

The determination of necrophagous entomofauna is a fundamental milestone for the determination of post mortem interval. This pioneer experimental design under laboratory conditions in Cameroon has enabled us to have an idea of arthropods that emerged from rearing maggots. Many necrophagous arthropods species invade rat carcasses for oviposition, larviposition and/or feeding. Their adults emerged along with some parasitoid wasps. These flies can be primarily considered as forensic entomology indicators. Further experiments will be performed to confirm these preliminary results in Central Africa, since the accuracy of Post Mortem Interval estimation is based on (1) many replicates of necrophagous insect studies, (2) effects of abiotic parameters environmental on insects development and (3) the quantity of energy used by necrophagous insects to complete their life cycle, which varies with temperature.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Catts E. Problems in estimating the post mortem interval in death investigations.

Journal of Agricultural Entomology. 1992; 9:245-255.

- Charabidzé D. Etude de la biologie des insects necrophages et application à l'expertise en entomologie medico-légale. Thèse de Doctorat/PhD. Université de Lille. 2008;2.
- Anderson GS, VanLaerhoven L. Initial studies on insect succession on carrion in Southwestern British Columbia. Journal of Forensic Sciences. 1996;41:617-625.
- Williams KA. Spatial and temporal occurrence of forensically important South African blowflies (Diptera: Calliphoridae).
 M. SC. Thesis, Rhodes University, South Africa; 2003.
- Feugang Youmessi FD. Etude des indicateurs entomologiques utilisables lors des enquêtes criminelles: cas des Arthropodes de Rattus Norvegicus (Berkenhout, 1769) Muridés souche Wistar à Yaoundé. Thèse de Doctorat Ph/D, Université de Yaoundé 1, Cameroun; 2014.
- Megnin JP, La faune des cadavres: Application de l'entomologie à la medecine légale. Paris: Masson et Gautheir-villars; 1894.
- Schoenly K, Reid W. Dynamics of heterotrophic succession in carrion arthropod assemblages: discrete seres or a continuum of change? Oecologia. 1987; 73:192–202.
- Boulton AJ, Lake PS. Dynamics of heterotrophic succession in carrion arthropod assemblages. A comment on Schoenly and Reid. Oecologia. 1988;76: 477-480.
- Bharti M, Singh D. Insect faunal succession on decaying rabbit in Punjabi, India. Journal of Forensic Sciences. 2003; 48:1-11.
- Villet MH. African carrion ecosystems and their insect communities in relation to forensic entomology. Pest Technology. 2011;5:1-15.
- 11. Dawson BM, Barton PS, Wallman JF. Contrasting insect activity and decomposition on pigs and Humans in an Australian environment: A preliminary study. Forensic Science International. 2020;316:1-8.
- Dautartas A, Kenyherez MW, Vidoli GM, Meadows JL, Mundorff A, Steadman DW. Differential decomposition among pig, rabbit and human remains. Journal of Forensic Science. 2018;63(6):1676-1683.

- Lutz L, Williams KA, Villet MH, Ekanem M. & Szpila K. Species identification of adult African Blowflies (Diptera: Calliphoridae) of forensic importance. International Journal of Legal Medecine. 2017;13:414-425.
- 14. Villet MH. History, accomplishments and challenges of forensic entomology in Africa in Tomberlin JK, Benbow ME. (eds). International dimensions and frontiers in forensic entomology. Taylor and Francis, Boca Raton. 2015;161-171.
- Prins AJ. Morphological and biological notes on six South African blowflies (Diptera: Calliphoridae) Commonly of forensic importance. Journal of Medical Entomology. 1982;48:738-752.
- Parry NJ, Mansell MW, Weldon CW. Seasonal, locality and habitat variation in assemblage of carrion associated Diptera in Gauteng province, South Africa. Journal of Medical Entomology. 2016;53:1322-1329.
- Ekenem MS, Dike JD. Arthropod succession on pig carcasses in South-Eastern Nigeria. Papéis Avulsos De Zoologia (São Paulo). 2010;50:561-570.
- Abajue MC, Ewuim SC, Akunne CE. Preliminary checklist of flies associates with pig carrions decomposing in Okya, Anambra State, Nigeria. Annual Research International. 2014;11(1):1899-1904.
- 19. Kurahashi H, Kirk-Spriggs A. The Calliphoridae of Namibia (Diptera: Oestroidea). Zootaxa. 2006;1322:1-131.
- 20. Tantawi TI, Wells JD, Greenberg B, El-Ghaffar HA. Arthropod succession on exposed rabbit carrion In Alexandria. Egyptian Journal of Medical Entomology. 1996;334:566-580.
- Tantawi TI, El-Kady EM, Greenberg B, El-Ghaffar HA. Fly larvae (Diptera: CAlliphoridae, Sarcophagidae, Muscidae) succession in rabbit carrion: variation observed in carcasses exposed at the same place. Journal of Egyptian German Society of Zoology. 1998;25:195-208.
- Aly SM, Jifang W, Xiang W, Jifeng C, Qinlai L, Ming Z. Identification of forensically important arthropods on exposed remains during summer season in northeastern Egypt. Journal of Central South University Medical Sciences. 2013; 38:1–6.
- 23. Ghaffar HAA, Moftah MZ, Favereaux A, Swidan M, Shalaby O. Mitochondrial DNAbased identification of developmental stages and empty puparia of forensically

important flies (Diptera) in Egypt. Journal of Forensic Science and Medicine. 2018; 4:7.

- Abd El-Gawad A, Badawy RM, Abd El-Bar MM, Kenawy MA. Successive waves of dipteran flies attracted to warfarinintoxicated rabbit carcasses in Cairo, Egypt. The Journal of Basic and Applied Zoology. 2019;80:56. Available:https://doi.org/10.1186/s41936-019- 0126-y.
- 25. Taleb M, Tail G, Kara FZ, Djedouani B, Toumi M. A preliminary study of the effects of environmental variables on early Diptera carrion colonizers in Algiers, Algeria. Journal of Entomology and Zoology Studies. 2015;4:181-188.
- Feugang Youmessi FD, Bilong Bilong CF, Cherix D, Djiéto-Lordon C. Biodiversity study of Arthropods collected on rat carrion in Yaounde, Cameroon: first study on forensic entomology in Central Africa. International Journal of Biosciences. 2012a;2(1):1-8.
- 27. Feugang Youmessi FD, De Coninck E, Hubrecht F, Braet Y, Bourguignon L, Bilong Bilong CF, Ateba Awona JC, Djiéto-Lordon C. First records on five species of Calliphoridae (Diptera) reared from maggot collected on rat carrions corpse during a forensic entomology experiment in the campus of the University of Yaounde I-Cameroon. International Journal of Biosciences. 2012b;2(3):75-80.
- Anderson GS. Factors that influence insect succession on carrion In: Byrd JH, Tomberlin JK, editors. Forensic entomology: The utility of arthropods in legal investigations. Third edition. Boca Raton (FL): CRC Press, Taylor and Francis; 2020.
- 29. Suchel F.G. Les climats du Cameroun, 2 tomes. Thèse de Doctorat d'Etat, Université Saint Etiènne (France). 1988;4.
- Kengne Fodouop, Atangana P. Une diversité du milieu biophysique exceptionnelle In Le Cameroun: Autopsie d'une exception plurielle en Afrique, Kengne Fodouop (Ed) –L'Harmattan. 2010;25-44.
- Anderson GS. Effects of Arson on forensic entomology evidence. Canadian Society of Forensic Science Journal. 2005;38:49-67.
- 32. Anderson GS, Cervenka VJ. Insects associated with the body/their use and analysis. In: Haglund WD, Sorg M, Editors, forensic entomology: the utility of arthropods in legal investigations, Boca

Raton FL. CRC Press. 2002;174-200.

- McAlpine JF, Peterson BV, Shewell GE, Teskey HJ, Vockeroth JR, Wood DM. Manual of Neartic Diptera.. Biosystematics Research Institute, Ottawa, Ontario. Research Branch Agricultural. 1981;1 (27):9-88.
- 34. Delvare G, Alberlenc HP. Les Insectes d'Afrique et d'Amérique. Clés pour la reconnaissance des familles.CIRAD, France (eds); 1989.
- Shaumar NF, Mohammad SK, Mohammad SA. Keys for identification of species of family Calliphoridae (Diptera) in Egypt. Journal of Egyptian Society of Parasitology. 1998;19(2):669-681.
- Carvalho CJBD. Muscidae (Diptera) of the Neotropical Region: Taxonomy. Editora UFPR, Brasil; 2006.
- Wyss C, Cherix D. Les insectes sur la scène de crime. Traité de l'entomologie Forensique. Deuxième Edition revue et augmentée. Eds: Presses polytechniques et universitaires romandes; 2013.
- Nihei SS, Carvalho CJBD. The muscini flies of the World (Diptera, Muscidae): identification key and generic diagnoses. Zootaxa. 2009;1976:1-24.
- Williams KA, Villet MH. Morphological identification of *Lucilia sericata, Lucilia cuprina* and their Hybrids (Diptera, Calliphoridae). Zookeys. 2014;420:69-85.
- 40. Kurahashi H, Chowanadisal L. Blow flies (Insects: Diptera: Calliphoridae) from Indochina. Species Diversity. 2001;6:185-242.
- 41. Rognes K, Paterson HEH. *Chrysomya chloropyga* (Wiedemann, 1818) and *C. putoria* (Wiedemann, 1830) (Diptera: Calliphoridae) are two different species. African enomology. 2005;23:56-61.
- 42. Couri MS. A key to the Afrotropical genera of Muscidae (Diptera). Revista Brasileira de Zoologia. 2007;24:175-184.
- 43. Lin Long I, Yang CH, Pai CY, Shiao SF. Population analysis of forensically important Calliphoridae On pig Corpses in Taiwan. Forensic Science International. 2010;9:25-34.
- Dajoz R. Les notions de biocénose et d'écosystème In Précis d'écologie, 5^e edition, Dunnod. 1985;248 276.
- 45. Monteiro-Filho ELA, Penereiro JL. A study on decomposition and succession on animal carcasses in an area of Sao Paulo, Brazil. Brazilian Journal of Biology. 1987; 47:289-295.

- 46. Velásquez Y. A checklist of arthropods associated with rat carrion in a mountain locality of Northern Venezuela. Forensic Science International. 2008;174:67-69.
- Warren JA, Anderson GS. A comparison of development times for *Protophormia terranovae* (Robineau Desvoidy) reared on different food substrates. Canadian Society of Forensic Sciences Journal. 2009;42:1616-171.
- 48. Simmons T, Adlam RE, Moffatt C. Debugging decomposition data-comp arative taphonomic studies and the difference of insects and carcass size on decomposition rate. Journal of Forensic Science. 2010;55(1):8-13.
- 49. Niederegger S, Pastuschek J, Mall G. Preliminary studies of the of the influence fluctuating temperature of on the development of various forensically relevant flies. Forensic Science International. 2020;199:72-78.
- 50. Connor M, Baigent C, Hansen ES. Testing the use of pigs as human proxies in decomposition studies. Journal of Forensic Science. 2018;63(5):1350-1355.
- Dao Hassane, Aboua LRN, Agboka Komi, Koffi AF, Djosso M. Influence des saisons sur l'activité des insects nécrophages dans le processus de decomposition des cadavres de proc (*Sus scrofa domesticus* L.) exposés à l'air libre en zone Subsoudanaise de Côte d'Ivoire. Afrique Science. French. 2019;15(1):361-376.
- Fuentes-López A, Ruiz C, Galián J, Romera E. Molecular identification of forensically important fly species in Spain using COI barcodes. Science & Justice. 2020;60:293-302.
- 53. Thyssen PJ, De Souza CM, Shimamoto PM, De Britto ST, Thiago CM. Rates of development of immatures of three species of *Chrysomya* (Diptera: Calliphoridae) reared in different types of animal tissues: implications for estimating the post mortem interval. Parasitology Research. 2014;113:3373-3380.
- 54. Whitaker AP. Development of blowflies (Diptera: Calliphoridae) on pig and human cadavers implications for forensic entomology casework. PHD thesis, king's college, London, UK; 2014.
- 55. Wang Y, Ma MY, Jiang XY, Wang JF, Li LL, Yin XJ, Wang M, Lai Y, Tao L.Y. Insect succession on remains of human and animals in Shenghen, China. Forensic Science International. 2017;271:75-86.

- Dawson BM, Wallman JF, Evans MJ, Barton PS. Is Resource Change a Useful Predictor of Carrion Insect Succession on Pigs and Humans? Geden C (Ed.). Journal of Medical Entomology; 2021. Available:tjab072.https://doi.org/10.1093/j me/tjab072
- Cervantès L, Dourel L, Gaudry E, Pasquereault T, Vincent B. Effect of low temperature in the development cycle of *Lucilia sericata* (Meigen) (Diptera: Calliphoridae): implications for the minimum post mortem interval estimation. Forensic Science Research. 2017;45:1-8.
- Lutz L, Mpreau G, Czuprynski S. An empirical comparison of decomposition and fly colonization of concealed carcasses in old and new world. International Journal of Legal Medecine. 2019;133(5):1593-1602.
- 59. Villet MH, Clitheroe Crystal, Williams KA. The temporal occurrence of flesh flies (Diptera, Sarcophagidae) at carrion-baited traps in Grahamstown, South Africa. African invertebrates. 2017;58(1):1-8.
- Williams KA, Villet MH. Spatial and seasonal distribution of forensically important blow flies (Diptera: Calliphoridae) in Makhanda, Eastern Cape, South Africa. Journal of Medical Entomology. 2019;56:1231-1238. Available:https://doi.org/10.1093/jme/tjz05 6
- 61. Kaneshrajah G, Turner B. *Calliphora vicina* larvae grow at different body tissue. International Journal of Legal Medecine. 2004;118:242-244.
- Clark K, Evans L, Wall R. Growth rates of the blowfly *Lucilia sericata*, on different body tissues. International Journal of Legal Medicine. 2006;156:145-149.
- Day DM, Wallmann JF. Influence of substrate tissue type on larval growth in *Calliphora augur* and *Lucilia cuprina* (Diptera: Calliphoridae). Journal of Forensic Science. 2006;51:657-663.
- 64. Ireland S, Turner B. The effect of larval crowding and food type on the size and development of the blowfly, *Calliphora vomitoria*. Forensic Science International. 2006;159:175-181.
- 65. Talebzadeh F, Ghadipasha M, Gharedaghi J, Yeksan N, Akbarzadeh K, Ali OM. Insect fauna of human cadavers in Tehran district. Journal of Arthropod-Borne Dis. 2017;11(3):363-370.
- 66. Maisonhaute JE, Forbes Shari L. Decomposition process and arthropods

succession on pig carcasses in Quebec (Canada). Canadian Society of Forensic Science Journal. 2020;10:1-26.

- 67. Da Silva MR, Moya BGE, De Carvalho Q MM. How photoperiods affect the immature development Of forensically important blowflies species *Chrysomya albiceps* (Calliphoridae). Parasitology Research. 2012;111:1067-1073.
- Kotteh M, Grieser J, Beck C. World map of the Köppen-Gieger Climate classification update. Metz. 2006;15:259-263.
- Matuszewski S, Szafalowicz M, Jarmusz M. Insects colonizing carcasses in an open and forest habitat in Central Europe: Search for indicators of corpse relocation. Forensic Science International. 2013;231: 234-239.
- Matuszewski S, Bajerlein D, Konwerski S, Szpila K. An initial study of insect succession and carrion decomposition in various forest habitats of Central Europe. Forensic Science International. 2018;180: 61-69.
- Fremdt H, Amendt J. Species composition of forensically blowflies (Diptera: Calliphoridae) and flesh flies (Diptera: Sarcophagidae) through space and time. Forensic Science International. 2014; 236:1-9.
- 72. Bernhardt V, Bàlint M, Verhoff MA, Amendt J. Species diversity and tissue specific dispersal of necrophagous Diptera on human bodies. Forensic Science, Medecine and Pathology. 2018;18: 1-8.
- 73. Harvey M, Gasz N, Woolley Z, Roberts L, Raven N, Colbart A, Law K, Marshall P, Voss S. Dipteran attraction to a variety of baits: implication for trapping studies as a tool for establishing seasonal presence of significant species. Journal of Medical Entomology. 2019;56(5):1283-1289.
- 74. Pitner S, Bugelli V, Weitgasser K, Zissler A, Sanit S, Luty L, Montidelli F, Campobasso CP, Steinbacher P, Amendt J. A field study to evaluate PMI estimation methods for advanced decomposition stages. International Journal of Legal Medecine. 2020;20:14-20.
- Richards CS, Williams KA, Villet MH. Predicting geographic distribution of seven blowflies species (Diptera: Calliphoridae) in South Africa. African Entomology. 2009a;17:170-182.
- 76. Richards CS, Price BW, Villet MH. Thermal ecophysiology of seven carrion-feeding blowflies in Southern Africa. Entomologia Experimentalis et Applicata.

2009b;131:11-19.

- Villet MH, Mackenzie B, Muller WJ. Larval development of the carrion-feeding flesh fly Sarcophaga (Liosarcophaga) tibialis Macquart (Diptera/ Sarcophagidae) at constant temperatures. African Entomology. 2006;14:357-366.
- Baz A, Cifrián B, Martin-Vega D. Patterns of Diversity and abundance of carrion insect assemblages in the Natural Park "Hoces del Rio Riaza" (Central Spain). Journal of Insect Science. 2014;14(162): 1-10.
- 79. Afravi Z, Sanei-Dehkordi A, Pirmohammadi M, Akbarzadeh K. Useful morphological characters of 3rd larval stages of three species of Sarcophagidae family 5diptera, Insecta. Journal of Entomology and Zoology Studies. 2015;3(5):483-486.
- Akbarzadeh K, Mirzaklanlou AA, Lotfalizadeh H, Malekian A, Hazratian T, Talarposhti RL, DarziBabapour R, Radi E, Afshaar AA. Natural parasitism associated sith species of Sarcophagidae family of Diptera in Iran. Annals of Tropical Medicine and Public Health. 2017;0:0-0.
- Akbarzadeh K, Saghafipour A, Jesri N, Karami-Jooshin M, Arzamani K, Hazratian T, Kordshouli RS, Afshaar AA. Spatial distribution of necrophagous flies of infraorder Muscomorpha in Iran Using Geographical Information System. Journal of Medical Entomology. 2018;55(5):1071-1085.
- Grassberger M, Franck C. Temperaturerelated development of the parasitoïd waps Nasonia vitripennis As forensic indicator. Medical and Veterinary Entomology. 2003;17:257-262.
- Turcheto M, Villemant-Aitlemkaden C, Vanin S. Two parasitoïds collected during an entomo-forensic investigation: The widespread Nasonia vitripennis (Hymenoptera: Pteromalidae) and the newly recorded Tachinaephagus zealandicus (Hymenoptera: Encyrtidae). Bolletino della Società Entomologica Italiana. 2003;135:108-115.
- Voss SC, Spafford H, Dadour IR. Hymenopteran parasitoïds of forensic importance: hostassociations, seasonality, and prevalence of parasitoïds of carrion flies in Western Australia. Journal of Medical Entomology. 2009;46:1210-1219.
- 85. Marchiori CH. Parasitoids of Chrysomya megacephala (Fabrcius) collected in

Itumbiara, Golas, Brazil. Review Soude publica. 2003;38:1-2.

86. Horenstein MB. & Salvo A. Community dynamics of carrion flies and their

parasitoïds in experimental carcasses in Central Argentina. Journal of Insect Science. 2012;12:1610.

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