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Evaluation of Thrombocytopenia: A Prospective Study at Sree Balaji Medical College and Hospital, India

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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Original Research Article

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ABSTRACT

Thrombocytopenia is a physiological deficiency in platelet counting. Fragmented RBC can be a chronic trigger for a subclinical micro angiopathy that results in chronic consumption of platelets. The platelet is a small, lentiform, anucleated cell that play a vital role in hemostasis and are produced in the bone marrow from megakaryocytes. To evaluate different etiological factors of thrombocytopenia by the study of clinical profile and laboratory parameters in patients with thrombocytopenia carried out in Sree Balaji Medical College and Hospital, Chennai. After evaluating all cases of thrombocytopenia, it is concluded that infective causes are more common than non-infective causes. Infections like dengue, malaria and septicemia were the common causes of thrombocytopenia along with megaloblastic anemia. Whenever thrombocytopenia is detected, a further investigation has to be done for specific diagnosis in the most of the cases so that appropriate treatment can be given.

Keywords: Fragmented RBC; thrombocytes; thrombocytopenia; megakaryocytes.

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1. INTRODUCTION

The definition of thrombocytopenia is platelet count < 1.5 lakhs/chum. It is the commonest platelet abnormality observed in clinical practice with different clinical expression. It may result from either decreased production or increased sequestration/destruction of platelets1. Destruction of platelets can be either immune or non-immune mediated. Numerous mechanisms may contribute in development of thrombocytopenia as seen in primary immune thrombocytopenia & hepatitis C virus infection. Thorough examination of the peripheral blood smear is the best way for narrowing the differential Megakaryocyte diagnosis. proliferation and platelet production are primarily regulated by interactions between thrombopoietin and its cell surface receptor, MPL [1-4]. Platelet production involves aggregation of components within the cell cytoplasm, segregation within a demarcation membrane system and organization into proplatelets [3]. The present study focuses on the evaluation of thrombocytopenia in different age groups who were established with thrombocytopenia. The platelet is a small, lentiform, anucleated cell14 that play a vital role in hemostasis and are produced in the bone marrow frommegakarvocytes. Mature megakaryocytes extend lona. branching processes. nominated proplatelets. which consists of platelet-sized swellings in tandem arrays that are connected by thin cytoplasmic bridges [4-7]. Red Blood Cells (RBC) on contact with prosthetic valves are continuously subject to damage. Fragmented RBC can be a chronic trigger for a subclinical micro angiopathy that results in chronic consumption of platelets.

Megakaryocytes arise from HSCs through a common megakaryocyte –erythroid progenitor cell that gives rise to erythroid precursors and megakaryoblasts. Megakaryocytes undertake endoreduplication as they mature, emanate in large cells (30-160 um). Maturation of megakaryocyte is dependent on transcription factors GATA1 and GATA 2 together with cofactor FOG1.The nuclei of great majority of normal polyploidy megakaryocytes form irregular lobes joined by strands of chromatin.

Infectious causes dominate in tropical countries like India. In congestive splenomegaly platelet sequestration occurs by redistribution of platelets from the circulatory pool to the splenic pool. Hemodilution is seen in patients who have received colloids, crystalloids & platelet poor blood products for massive hemorrhage [8,9]. Platelets escaped identification for a long time, because of their small size and the limited resolution of early microscopes, in 1735, the German physician and poet Paul Gottlieb Werlhof provided the first detailed description of 'morbus maculosus haemorrhagicus' now known as immune thrombocytopenia (ITP), these blood cells were unknown [10,11]. The discovery of platelets had to wait until 1882, when the Italian pathologist Giulio Bizzozzero, described in detail these small elements and the relationship between platelet adhesion and aggregation, fibrin formation and deposition [12].

One year after the brilliant insight of Bizzozero, Brohm identified the link between thrombocytopenia and ITP [6]. The intuition of Kaznelson, in 1916, that the spleen was responsible for platelet destruction led to the identification of splenectomy as a potent treatment for this disease [13,14,15]. The present study aimed to nalyse the associated causes for the development of thrombocytopenia in patients admitted to Sree Balaji Medical college and Hospital, Chennai.

2. MATERIALS AND METHODS

This prospective study was conducted in the Sree Balaji medical college and hospital from march 2017 to October 2018. This study included 100 subjects who presented to the hematology department and medical OP departments of Sree Balaji Medical College and hospital. Ref. No. 002/SBMC/IHEC/2017/869.

2.1 Inclusion Criteria

Patients presenting to the hematology department and medical OP departments who were found to have thrombocytopenia, with platelet count less than 150 x 109/L in whom complete clinical and laboratory parameters were available.

2.2 Exclusion Criteria

Patients with platelet count more than 150 x 109/L, patients presented with massive hemorrhage, and who received massive colloid or crystalloid transfusion for volume loss are not included in our study. A detailed clinical history was taken. General and systemic examination was done in each patient who were included in the study population. Provisional diagnosis was made based on clinical examination and

laboratory parameters wherever necessary.

Peripheral venous blood was collected from antecubital vein. Appropriate amounts of blood were transferred into sodium citrate 3.2 % for coagulation estimation of profile, and tripotassium EDTA vacutainer for complete blood count. For biochemical analysis such as total leucocyte count, differential count, haemoglobin, hematocrit, Mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), red cell distribution width, platelet count, mean platelet volume and platelet distribution width. erythrocyte sedimentation rate (ESR) was measured by vesmatic cube 30 and Westergren's method. For PT, aPTT and fibrinogen estimation, blood with appropriate amount of 3.2 % sodium citrate was used. PT, aPTT and fibrinogen was assessed by using coagulatory method in BS 390 fully automatic analyzer. D-dimer levels were also estimated in They were stained by selected cases. Leishman's stain. For hemoparasite visualization. Giemsa stain was used. Bone marrow aspirate was taken from posterosuperior iliaccrest with help of 16G bone marrow aspiration needle. Smears were stained with Leishman's stain. Bone marrow trephine biopsy was performed in applicable cases and hematoxylin and eosin (H & E) stained paraffin sections were examined.

3. RESULTS AND DISCUSSION

In the present study maximum number of patients were in the age group 20-39 years (42 cases, ie.,42 %). (Table.1, Fig. 1).

In our study maximum number of patients presenting with thrombocytopenia were males (68%). (Table 2, Fig. 2).

The total number of our study population were analysed and categorized according to the final diagnosis. They distributed as follows. (Table .3, Fig. 3).

3.1 Diagnosis Associated with Thrombocytopenia

The most common cause observed was dengue 29% followed by malaria (15%), septicemia (13%), megaloblastic anemia (9%), liver disorder (9%), leukemia (7%), HIV (human immunodeficiency virus) infection (4%), ITP (3%) Drug induced thrombocytopenia (3%), DIC (2%), Tuberculosis (2%), Aplastic anemia (2%), MDS (1%), Hypersplenism (1%).

These cases were then analysed with the entire clinical andlaboratory profiles and the possible pathogenesis outlined. (Table 4, Fig. 4).

Infective causes were found to be the most common causes of thrombocytopenia in our study.

In our study the most common cause observed was dengue 29% followed by malaria (15%), septicemia (13%), megaloblastic anemia (9%).Khatib et al. reported 10% Dengue, 29.67% malaria, 12.67% septicemia, and 15.67% megaloblastic anemia cases [16].

Kumar et al. [17] reported 32.63% of malaria, 15.78% dengue and 31.57% septicemia cases in their study [17]. Nair et al. reported 14% dengue, 9% malaria, 26% of septicemia cases in their patients with thrombocytopenia [18]. Patil et al. reported 15% dengue, 54% malaria and 4% septicemia cases [8]. Gutthi et al. reported 35% of malaria, 5% septicemia and 34% dengue cases in their study [19]. Study by paramjit et al. reported malaria 57.7%, dengue 27.7% and septicemia 4.7% cases [20]. Sanjay et al. reported dengue 30%, malaria 12.5% and megaloblastic anemia 21.6% in their study Yadav et al. reported dengue 26.8%, malaria 24.4%, and septicemia 4.4% in their studv [21,22,23,24,25].

Table 1. Age distribution of cases of	f thromboc ^y	ytopenia
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Age (years)	No. of cases	Percentage	
0 – 19	28	28 %	
20 - 39	42	42%	
40 – 59	22	22%	
60 – 79	8	8%	
Total	100	100%	



Fig. 1. Age distribution of cases of thrombocytopenia

Table 2. Sex distribution of	of cases of	f thrombocytopenia
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Sex	No. of cases	percentage
Male	68	68%
Female	32	32%
Total	100	100%



Fig. 2. Sex Distribution of cases of thrombocytopenia

S.no	Diagnosis	No. of cases %
1	Dengue	29 (29%)
2	Malaria	15 (15%)
3	Septicemia	13 (13%)
4	Megaloblastic anemia	9 (9%)
5	Liver disorder	9 (9%)
6	Leukemia	7 (7%)
7	HIV infection	4 (4%)
8	ITP	3 (3%)
9	Drug induced	3 (3%)
10	DIC	2 (2%)
11	Tuberculosis	2 (2%)
12	Aplastic anemia	2 (2%)
13	MDS	1 (1%)
14	Hypersplenism	1 (1%)
	Total	100 (100%)

Table 3. Diagnosis associated with thrombocytopenia



Fig. 3. Diagnosis associated with thrombocytopenia

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S.no	Etiological factor	No of cases (%)
1	Decreased production	9 (9%)
2	Ineffective haematopoiesis	10 (10%)
	Congestive splenomegaly with	
3	hypersplenism	1 (1%)
4	Infective causes	63(63%)
5	Increased peripheral destruction	8 (8%)
6	Liver disorders	9 (9%)
	Total	100 (100%)



Fig. 4. Pathogenesis based categorization of thrombocytopenia

Our study included 2 cases of acute myeloid leukemia (AML) (one AML-M4) and 1 case of ALL. AML- M4 in peripheral smear showed increased number of both myeloblasts and monoblastsalong with reduced number of platelets (Fig. 1) with hypercellular marrow, heterogenous cells, including immature monocytes and neutrophils. Peripheral smear of ALL showed leukoerythroblastosis, occasional reactive lymphocytes with thrombocytopenia. Bone marrowwas hypercellular with infiltration by 90% lymphoblasts.

The two cases of Chronic myelogenous leukemia (CML) and one Chronic myelomonocytic leukemia (CMML) case were included in our study. Peripheral smear of CMML shows leucoerythroblastic picture with 15% myeloblasts (Fig. 2) and thrombocytopenia, bone marrow showed 43% myeloblasts, with evidence of

hemophagocytosis. One case of Chronic lymphocytic leukemia (CLL) with peripheral smear showed thrombocytopenia with 63% lymphocytes, 32% neutrophils, monocytes 3%. (Fig. 3) Bone marrow biopsy showed nodular infiltration oflymphocytes. 2 cases of aplastic anaemia were included in our study, bone marrow aspiration showed hypocellular smear with a relative pancytopenia of normal hematopoietic cells along with scattered lymphoplasmacytic infiltration (Figs. 4 & 5). 9 cases of Megaloblastic anaemia were included in our study.We had one case of primary MDS in study. Peripheral smear showed our pancytopenia with leucoerythroblastic picture, macrocytes, macro - ovalocytes, nucleated RBCs with dysplasias and 4% blasts. bone marrow showed erythroid hyperplasia (70%) with dysplasia particularly in erythroid series. (Fig. 6).



Fig. 5. Photomicrograph of peripheral blood smear showing monoblasts with abundant vacuolated cytoplasm and prominent nucleoli background showing reduced number of platelets in a case of AML-M4 (Leishman 100x)

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Fig. 6. Photomicrograph of peripheral blood smear in a case of CMML showing myeloblasts and monocytes (Leishman 100x)



Fig. 7. Photomicrograph of peripheral blood smear showing increased number of mature lymphocytes and smudge cells background showing reduced number of platelets in a case of CLL (Leishman 100x)



Fig. 8. Photomicrograph of hypocellular bone marrow aspiration smear showing lymphoplasmacytic infiltrate and mast cells in a case of Aplastic anemia.(Leishman100x)

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Fig. 9. Photomicrograph of hypocellular bone marrow biopsy showing15% cellularity in same case of Aplastic anemia (H & E 10x)



Fig. 10. photomicrograph of bone marrow aspiration smear showing dyserythropoiesis in a case of MDS. (Leishman 100x)



Fig. 11. Photomicrograph of peripheral blood smear showing Schizont phase of plasmodium vivax in a patient with malaria, background shows decreased number of platelets. (Leishman 100x)



Fig. 12. Photomicrograph of bone marrow aspiration smear showing clusters of micromegakaryocytes in a case of ITP (Leishman 100x)

3.2 Liver Disorders

We had total 9 cases of hepatic dysfunction in our study.

CONCLUSION

Males were more commonly affected with thrombocytopenia than females in our study. Age group between 20-39 years people were more commonly affected than other age groups in our study, in most of the other studies also prevalence of thrombocytopenia is more common in this age group.

CONSENT

As per international standard or university standard, patients' written consent has been collected and preserved by the author(s).

ETHICAL APPROVAL

The study was approved by the Institutional Ethics Committee.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Evans RS, Takahashi K, Duane RT, Payne R, Liu CK. Primary thrombocytopenic purpura and acquired hemolytic anemia: evidence for a common etiology. AMA archives of internal medicine. 1951;87(1):48-65.
- Schick BP. Hope for treatment of thrombocytopenia. New England Journal of Medicine. 1994;331(13):875-6.
- 3. Bain BJ, Clark DM, Lampert IA, Wilkins BS. Bone marrow pathology. John Wiley & Sons. 2008.
- 4. Deutsch VR, Tomer A. Megakaryocyte development and platelet production. British journal of haematology. 2006;134(5):453-66.
- 5. Choudhary PK, Sing SK, Basnet RB. Study of megakaryocytes in bone marrow aspiration smears in patients with thrombocytopenia. Journal of Pathology of Nepal. 2013;3(6):476-81.
- 6. Holinstat M. Normal platelet function. Cancer and Metastasis Reviews. 2017;36(2):195-198.
- Thon JN, Italiano JE. Platelet formation. In Seminars in hematology. WB Saunders. 2010:47(3):220-226.
- 8. Patil P, Solanke P, Harshe G. To study clinical evaluation and outcome of patients with febrile thrombocytopenia. Int J Sci Res Publications. 2014;4(10):01-3.
- 9. Aster RH. Pooling of platelets in the spleen: Role in the pathogenesis of. The Journal of clinical investigation. 1966;45(5):645-57.
- 10. Rodgers GM. Thrombocytopenia: Pathophysiology and classification.

InWintrobe's Clinical Hematology: Thirteenth Edition 2013 Oct 17. Wolters Kluwer Health Adis (ESP).

- 11. Werlhof PG. Philologa inquiry doctor and some of the statues or coal. Impensis Nicolai Foersteri and child; 1735.
- 12. Bizzozero J. On a new constituent of the blood and its role in thrombosis and blood clotting. Archive for pathological anatomy and physiology and for clinical medicine. 1882;90(2):261-332.
- 13. Balduini CL, Melazzini F. Research at the heart of hematology: Thrombocytopenias and platelet function disorders; 2017.
- 14. Kaznelson P. Disappearance of hemorrhagic diathesis in a case of essential thrombocytopenia (frank) after spleen excitement: splenogenic thrombolytic purpura. Vienna KlinWochenschr. 1916; 29:1451st.
- Harrington WJ, Minnich V, Hollingsworth JW, Moore CV. Demonstration of a thrombocytopenic factor in the blood of patients with thrombocytopenic purpura. The Journal of laboratory and clinical medicine. 1951;38(1):1-0.
- 16. Yasmeen Khatib, Vaishali Jain, Richa Patel. One year study of thrombocytopenia in a peripheral hospital of Mumbai. 2016;6(4):26-30.
- Kumar P, Chandra K. A clinical study of febrile thrombocytopenia: A hospital-based retrospective study. Indian Journal of Clinical Practice. 2014;24(10):952-7.
- 18. Nair PS, Jain A, Khanduri U, Kumar V. A

study of fever associated with thrombocytopenia. JAPI. 2003;51:1173.

- Gutthi LP, Vegesna S, Pundarikaksha V, Kolla S, Gundapaneni M, Karimi PK. A study of clinical and lab profile of fever with thrombocytopenia. Internat J Contempt Medic Res. 2017;4(5):1057- 61.
- 20. Paramjit E, Rao R, Sudhamani S, Roplekar P, Shaffi Z, Roy S. Spectrum of thrombocytopenia: A clinicopathological study with review of the literature. Muller Journal of Medical Sciences and Research. 2016;7(2):121.
- Yadav V, Singhai A. Study of febrile thrombocytopenia in Malwa region of India. Asian Journal of Medical Sciences. 2017;8(5):83-6.
- 22. Bain BJ, Bates I, Laffan MA. Dacie and lewis practical haematology e-book. Elsevier Health Sciences; 2016.
- 23. Microkrom Microscopystains Microkrom. Basic Guidelines for Quality Staining with Romanowsky Stains Coral Clinical Systems.
- 24. Patne SV, Chintale KN. Clinical profile of patients with thrombocytopenia at tertiary health care centre. International Journal of Advances in Medicine. 2017;4(6): 1551-6.
- Yamaguchi S, Kubota T, Yamagishi T, Okamoto K, Izumi T, Takada M, et al. Severe thrombocytopenia suggesting immunological mechanisms in two cases of vivax malaria. American journal of hematology. 1997;56(3):183-6.

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